SESQUITERPENE DERIVATIVES FROM *OTHONNA* AND RELATED SPECIES

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Key Word Index—Othonna species; Senecio macrospermus; Euryops pedunculatus; Compositae; sesquiterpenes; caryophyllene derivative; bicyclogermacrene derivative; furoeremophilanes; bisabolones.

Abstract—The investigation of several South African representatives of the tribe Senecioneae afforded in addition to known compounds a caryophyllene and a bicyclogermacrene derivative, two furoeremophilanes, three bisabolone derivatives and a guaiane derivative.

INTRODUCTION

From the large South African genus Othonna with about 150 species so far 20 species have been studied chemically [1-8]. Most species afforded furanoeremophilanes which are also widespread in Senecio and Euryops. Only a few species gave other constituents like diterpenes [4], phydroxyacetophenones [1, 8] and different sesquiterpenes [1]. We now have studied seven further species and a South African Senecio and a Euryops species. The results are presented in this paper.

RESULTS AND DISCUSSION

The aerial parts of Othonna leptodactyla Harv. afforded in addition to known furoeremophilanes (Table 2) a new diol (1: $R = CH_2OH$, $R^1 = H$, $R^2 = OAng$, $R^3 = H$, $X = H_2$), the structure of which followed from its ¹H NMR spectrum which was close to that of similar compounds from Othonna species [2]. The natures and the relative positions of the oxygen functions were also deduced from the ¹H NMR data. Spin decoupling allowed the assignment of all signals though a few were multiplets. The ¹³C NMR spectrum also supported the structure.

The aerial parts of O. obtusiloba (DC) Sch. Bip. gave several known furoeremophilanes (Table 2) and one new ester (1: R = Me, $R^1 = iVal$, $R^2 = H$, $X = H_2$, $R^3 = OH$). The structure could be deduced easily from the ¹H NMR spectrum which was close to that of the corresponding senecioate [9]. The nature of the ester group followed from the typical signals. Furthermore the pmethoxyphenethyl alcohol esters 9 and 10 were present. Their structures also followed from the spectroscopic data

The aerial parts of O. cf. graveolens O. Hoffm. gave no furoeremophilanes but two sesquiterpene senecioates (11 and 12). The structure of 11 was deduced from the molecular formula $(C_{20}H_{30}O_2)$ and the ¹H NMR data. The typical signals of a senecioate indicated the nature of the oxygen function. The remaining signals were close to those of caryophyllene. The position of the ester was determined by NOE difference spectroscopy which gave clear effects between H-13 and H-9 as well as between

H-14 and H-1. The corresponding 13-O-acetate has been isolated from a *Helichrysum* species [10].

The ¹H NMR spectrum of 12 clearly showed that we were dealing with a senecioyloxy derivative of bicycloger-macrene. Accordingly, most signals were close to those of the hydrocarbon, only one methyl singlet being replaced by a pair of doublets (H-12). Again the position of the ester group was determined by clear NOEs between H-13, H-6 and H-7. Spin decoupling allowed the assignment of all signals. The aerial parts of four further *Othonna* species gave known compounds (Table 2).

The reinvestigation of the aerial parts of Euryops pedunculatus N.E.Br. gave in addition to compounds

Table 1. ¹H NMR spectral data of 5-7 (400 MHz, CDCl₃, TMS as int. standard)

	5	6	7	
H-1	5.75	5.85	5.75 d	
H-4	3.42	3.43	3.41 d	
Η-5α	2.56	2.59	2.54 ddd	
Η-5β	2.19	2.19	2.18 dd	
H-6	2.77	2.76	2.77 ddd	
H-8	5.07	5.09	5.05 m	
H-9	2.34	2.32	2.32 m	
H-10	5.03	5.04	5.02 br t	
H-12	1.65	1.64	1.65 br s	
H-13	1.59	1.59	1.59 br s	
H-14	5.10	5.07	5.08 br s	
H-14'	5.23	5.22	5.22 br s	
H-15	1.42	1.44	1.43 s	
OAng	6.09	6.04	qq	
	1.97	1.95	— dq	
	1.87	1.87	dq	
OSen	5.63	5.64	5.68, 5.64 qq	
	2.12	2.14	2.18, 2.14 d	
	1.84	1.84	1.84, 1.88 d	

J (Hz): 1,6 = 12; 4,5 = 4.5; $5\alpha,5\beta = 16$; $5\alpha,6 = 7.5$; $5\beta,6 = 11$; 9,10 = 7; OAng: 3',4' = 7; 3',5' = 4',5' = 1.5; OSen: 2',4' = 2',5' = 1.3.

Table 2. Investigated species and constituents

Name (voucher)	Wt. of aerial parts (g)	Constituents	
Othonna cf. arbuscula (Thunb.) Sch. Bip. (81/155)			
O. furcata (Lindl.) Bruce (81/157)	30	$5 \text{ mg 1 } (R = Me, R^1 = Sen, R^2 = OSen, R^3 = H, X = O) [5], 7 \text{ mg 1 } (R = Me, R^1 = Meacr, R^2 = OSen, R^3 = H, X = O) [5], 9 \text{ mg 1 } (R = Me, R^1 = iBu, R^2 = OMeacr, R^3 = H, X = O) [4], 2 \text{ mg 1 } (R = Me, R^1 = Meacr, R^2 = OTigl, R^3 = H, X = O) [5]$	
O. cf. graveolens O. Hoffm. (81/156)	30	11 mg 11, 7 mg 12	
O. lasiocarpa (DC) Sch. Bip. (81/161)	22	11 mg 1 (R = Me, R^1 = Sen, R^2 = OSen, R^3 = H, X = O), 15 mg 1 (R = Me, R^1 = Meacr, R^2 = OSen, R^3 = H, X = O), 17 mg 1 (R = Me, R^1 = iBu, R^2 = OMeacr, R^3 = H, X = O), 5 mg 1 (R = Me, R^1 = Meacr, R^2 = OTigl, R^3 = H X = O), 11 mg 2	
O. leptodactyla Harv. (81/170)	160	18 mg 1 (R = CH ₂ OH, R ¹ = H, R ² = OAng, R ³ = X = H), 10 mg 1 (R = CO ₂ H R ¹ = iBu, R ² = OAng, R ³ = X = H) [4], 10 mg 1 (R = CO ₂ H, R ¹ = iVal, R ² = OAng, R ³ = X = H) [4]	
O. obtusiloba (DC) Sch. Bip. (81/162)	75	800 mg 1 (R = Me, R ¹ = Meacr, Sen, R ² = R ³ = H, X = O), 8 mg 1 (R = Me, R ¹ = Sen, R ² = X = H, R ³ = OH) [10], 2 mg 9, 2 mg 10, 4 mg cis- and 2 mg transcaryophyllene	
O. sedifolia DC (81/166)	110	80 mg ent-kaurenic acid, 10 mg 16,17-epoxy-ent-kaurenic acid, 4 mg manoyl oxide, 3.5 mg 17-hydroxyisokaurenic acid	
Euryops pedunculatus N.E.Br. (81/251) collected in Transvaal	100	3 mg 1 ($R = Me$, $R^1 = Ac$, $R^2 = R^3 = H$, $X = O$), 3 mg of the corresponding 1,10-dehydro and 3 mg of the 1,10-epoxy derivative [13], 3 mg 8	
Senecio macrospermus DC (Hilliard 3/3/82) collected in Natal	250	42 mg 3 (R = OAc) [18], 12 mg 3 (R = OH) [19], 40 mg 3 (R = H) [19], 15 mg 4 (R = H) [20], 30 mg 4 (R = OMe), 12 mg 5, 7 mg 6, 16 mg 7, 7 mg 5 (R = Sen, R ¹ = H), 3 mg 5 (R = Ang, R ¹ = OAng)	

isolated previously [11] (Table 2) the guaiane derivative 8. The structure followed from the 1 H NMR spectrum. Starting with the olefinic methyl spin decoupling, the whole sequence could be determined. From the observed couplings of H-1, the relative configuration at C-1 and C-10 could be deduced by inspection of models. As usual all sesquiterpenes from Compositae have the 7α -H configuration, also H-1 and H-10 are most likely α -oriented.

The aerial parts of Senecio macrospermus DC afforded several furanoeremophilanes of the cacalol type and several bisabolone derivatives (Table 2). In addition to known ones [12, 13], three new bis-esters (5-7) were isolated. The ¹H NMR spectra (Table 1) were of course close to that of the bisangelate [12]. Only the chemical shifts of H-1 and H-8 showed small differences. As in similar cases the replacement of an angelate by a seneciote residue caused a small upfield shift of the corresponding proton under the ester group. The configuration at C-8 could not be determined.

The new results on the chemistry of Othonna species established again that most species contain furoeremophilanes. But again one species had diterpenes while another one afforded only monosubstituted sesquiterpenes. There is still no clear chemical differentiation of the genera Othonna, Euryops and Senecio. In all three cases furoeremophilanes are most widespread, and there is a tendency in Othonna species to accumulate compounds with oxygen functions at C-15. However, there are several groups where these compounds are lacking [14]. Clearly our knowledge is still too limited for any final conclusions to

be drawn, especially as the taxonomy of the whole tribe Senecioneae is still not solved.

EXPERIMENTAL

The Othonna species were grown in a greenhouse of the Botanic Research Institute in Pretoria. The air-dried aerial parts were extracted and from the extracts obtained the separation of the constituents was achieved as reported previously [15]. The results are summarized in Table 2. The conditions of the isolation of new compounds are given in parentheses. Known compounds were identified by comparing their 400 MHz ¹H NMR spectra with those of authentic material.

3β-Angeloyloxy-6β,15-dihydroxy-10βH-furoeremophilane. Colourless oil; $IR v_{max}^{\rm CCL}$ cm⁻¹: 3450 (OH), 1720, 1650 (C=CCO₂R); MS m/z (rel. int.): 330.183 [M - H₂O]⁺ (40) (calc. for C₂₀H₂₆O₄:330.183), 230 [330 - RCO₂H]⁺ (24), 83 [C₄H₇CO]⁺ (100), 55 [85 - CO]⁺ (82); ¹H NMR (CDCl₃); δ2.16 (m, H-1), 1.41 (brd, H-1'), 1.90 (m, H-2), 5.30 (ddd, H-3), 2.26 (ddd, H-4), 4.57 (br s, H-6), 2.56 (m, H-9), 7.00 (br s, H-12), 1.99 (d, H-13), 1.35 (s, H-14), 3.92 and 3.67 (dd, H-15); OAng: 6.05 qq, 1.98 dq, 1.78 dq; J (Hz): 1,1' = 15; 3,4 = 5; 4,15 = 7.5; 4,15' = 11; 12,13 = 1; 15,15' = 12; OAng: 3',4' = 7; 3',5' = 4',5' = 1.5; ¹³C NMR (CDCl₃, C-1-C-15): 25.6 t, 22.9 t*, 81.1 d, 41.8 s, 67.3 d, 120.1 s, 148.3 s, 22.7 t*, 37.9 d, 119.0 s, 137.9 d, 8.3 q, 22.6 q, 67.0 t; OAng: 167.3 s, 127.6 s, 138.4 d, 20.7 q, 15.6 q (*values may be interchangeable); [α] $\frac{24^{*}}{5}$ 5 (CHCl₃; c 0.9) (PTLC: Et₂O-petrol, 3:7, R_f 0.6).

 6β -Isovaleryloxy-10 β -hydroxyfuroeremophilane. Colourless oil; IR $\nu \frac{\text{CGL}}{\text{cm}^{-1}}$: 3570 (OH), 1720, 1650 (C=CCO₂R); MS m/z

(rel. int.): 334.214 [M] * (1.4) (calc. for $C_{20}H_{30}O_4$: 334.214), 232 [M - RCO₂H] * (28), 85 [C₄H₉CO] * (45), 57 [85 - CO] * (100);

¹H NMR (CDCl₃): 6.19 (s, H-6), 3.20 and 2.66 (brd, H-9), 7.10 (brs, H-12), 1.91 (d, H-13), 1.02 (s, H-14), 0.84 (d, H-15); OiVal: 2.19 (d, H-2'), 2.08 (m H-3'), 0.91 (d, 6H); J (Hz): 4,15 = 7; 9,9' = 18; 12,13 = 1; 2',3' = 3',4' = 3',5' = 7; (PTLC: Et₂O-petrol, 1:9, two developments, R_f 0.45).

1α-Senecioyloxy-8-angeloyloxy-3β,4β-epoxybisabola-7(14),10-dien-2-one (5). Colourless oil; $IR v \stackrel{CCQ}{\max} cm^{-1}$: 1730 (C=O, C=CO₂R), 1660 (C=C); MS m/z (rel. int.): 430.236 [M]⁺ (0.4) (calc. for C₂₅H₃₄O₆: 430.236), 361 [M - C₅H₉]⁺ (5), 330 [M - RCO₂H]⁺ (5), 230 [330 - RCO₂H]⁺ (12), 83 [C₄H₇CO]⁺ (100); [α] $\frac{7}{100}$ -40 (CHCl₃; c0.81) (HPLC: RP8, MeOH-H₂O, 4:1, R, 4.5 min).

1α-Angeloyloxy-8-senecioyloxy-3 β ,4 β -epoxybisabola-7(14),10-dien-2-one (6). Colourless oil; IR ν_{max}^{CCl} cm⁻¹: 1730 (C=CCO₂R, C=O), 1660 (C=C); MS m/z (rel. int.): 430.236 [M]⁺ (0.5) (calc. for

 $C_{25}H_{34}O_6$: 430.236), 361 [M-C₅H₉]⁺ (6), 330 [M-RCO₂H]⁺ (1), 230 [330-RCO₂H]⁺ (3), 83 [C₄H₇CO]⁺ (100); [α] $\frac{1}{2}$ ⁰ - 58 (CHCl₃; c 0.37) (HPLC: RP 8, MeOH-H₂O, 4:1, R, 5.0 min).

1α,8-Bissenecioyloxy-3β,4β-epoxybisabola-7(14),10-dien-2-one (7). Colourless oil; IR $v_{\text{max}}^{\text{CCL}_4}$ cm $^{-1}$: 1730 (C=O, C=CCO₂R), 1660 (C=C); MS m/z (rel. int.): 430.236 [M] $^+$ (0.5) (calc. for C₂₅H₃₄O₆: 430.236), 361 [M - C₅H₉] $^+$ (6), 330 [M - RCO₂H] $^+$ (4), 230 [330 - RCO₂H] $^+$ (6); [α] $^{26}_{\text{c}}$ - 75 (CHCl₃; c 0.62) (PTLC: CH₂Cl₂-C₆H₆-Et₂O, 4:4:1, R_f 0.65).

3-Oxo-gual-4-en-11-ol (8). Colourless oil; IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3600 (OH), 1685, 1628 (C=CC=O); MS m/z (rel. int.); 218.167 [M - H₂O] + (45) (calc. for C₁₅H₂₂O: 218.167), 203 [218 - Me] + (6), 178 [M - O=CMe₂] + (11), 59 [Me₂COH] + (100); CIMS m/z (rel. int.); 237 [M + 1] + (100), 219 [237 - H₂O] + (44); ¹H NMR (CDCl₃); δ 3.04 (br d, H-1), 2.47 (dd, H-2), 1.96 (d, H-2'), 2.99 (br d, H-6), 2.10 (br dd, H-6'), 1.62 (m, H-7), 2.00 (m, H-10), 1.19

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(s, H-12), 1.15 (s, H-13), 0.53 (d, H-14), 1.60 (brs, H-15) [J(Hz): $1,2=7; 1,10\sim 2; 2,2'=19; 6,6'=20; 6.7=13; 6.15\sim 1.5; 10,14=7]; [\alpha]_{D}^{24^{\circ}}+18 (CHCl_{3}, c 0.2) (PTLC: Et₂O-MeOH, 20:1, <math>R_{f}$ 0.35).

2-[4-Methoxyphenyl]-ethylsenecioate (9). Colourless oil; $IR \ v_{\max}^{CC4} \ cm^{-1}$: 1720 (C=CCO₂R); MS m/z (rel. int.): 234.126 [M] + (0.4) (calc. for $C_{14}H_{18}O_3$: 234.126), 134 [M - RCO₂H] + (100), 83 [C₄H₇CO] + (26); ¹H NMR (CDCl₃): δ 7.15 (d, H-2, H-6), 6.84 (d, H-3, H-5), 2.90 (t, H-7), 4.26 (t, H-8); OSen: 5.67 qq, 2.15 and 1.88 d [J (Hz): 2,3 = 8; 7,8 = 7; OSen: 2',4' = 2',5 = 1] (PTLC: Et₂O-petrol, 1:9, R_f 0.70).

2-[4-Methoxyphenyl]-ethylisovalerate (10). Colourless oil; $\text{IR v}_{\text{max}}^{\text{CCI}_4} \text{ cm}^{-1}$: 1740 (CO₂R); MS m/z (rel. int.): 236.141 [M] + (25) (calc. for C₁₄H₂₀O₃: 236.141), 134 [M - RCO₂H] + (100),

1H NMR (CDCl₃): δ 7,13 (d, H-2, H-6), 6.84 (d, H-3, H-5), 2.87 (t, H-7), 4.24 (t, H-8); OiVal: 2.17 d, 2.07 tqq, 0.90 d [J (Hz): 2,3 = 8; 7,8 = 7; OiVal: 2',3' = 3',4' = 3',5' = 7] (PTLC: Et₂O-petrol, 1:9, R_f 0.68).

13-Senecioyloxy-caryophyllene (11). Colourless oil; $IR \, v_{\rm max}^{\rm CHCl_3} \, cm^{-1}$: 1715 (C=CCO₂R); MS m/z (rel. int.): 302.225 [M] + (0.7) (calc. for C₂₀H₃₀O₂: 302.225), 202 [M - RCO₂H] + (5.5), 83 [C₄H₇CO] + (100); ¹H NMR (CDCl₃): δ 1.61 (t, H-1), 5.27 ($br \, dd$, H-5), 2.35 ($br \, q$, H-9), 4.99 and 4.86 ($br \, s$, H-12), 4.15 and 4.05 (d, H-13), 1.12 (s, H-14), 1.61 ($br \, s$, H-15); OSen: 5.69 qq, 2.17 and 1.90 d [d (Hz): 1,2 = 1,9 = 9; 5,6 = 12; 5,6' = 4; 9,10 = 10; 13,13' = 11] (PTLC: CH₂Cl₂-C₆H₆, 1:1, two developments, R_f 0.55).

12-Senecioyloxy-bicyclogermacrene (12). Colourless oil; $IR v_{max}^{CHCl_3} cm^{-1}$: 1715, 1650 (C=CCO₂R); MS m/z (rel. int.): 302.225 [M] $^+$ (0.6) (calc. for $C_{20}H_{30}O_2$: 302.225), 202 [M $^-$ RCO₂H] $^+$ (7.5), 187 [202 $^-$ Me] $^+$ (9), 83 [C₄H₇CO] $^+$ (100), 55 [83 $^-$ CO] $^+$ (35); 1 H NMR (CDCl₃): 4.84 (br dd, H-1), 2.03 (m, H-2 α), 2.10 (br d, H-2 β), 1.85 (m, H-3 α), 2.21 (ddd, H-3 β), 4.40 (br d, H-5), 1.49 (dd, H-6), 0.82 (ddd, H-7), 2.03 (m, H-8 α), 1.31 (dddd, H-8 β), 1.73 (ddd, H-9 α), 2.43 (ddd, H-9 β), 4.22 and 4.14 (d, H-12), 1.19 (s, H-13), 1.48 (d, H-14), 1.67 (d, H-15); OSen: 5.72 qq, 2.17 and 1.90 d [J (Hz): 1,2 α = 6; 1,2 β = 2 α ,2 β = 3 α ,3 β = 12; 2 α ,3 β = 2 β ,3 β = 3; 5,6 = 7,8 β = 11; 6,7 = 8; 7,8 α = 5; 8 α ,8 β = 9 α ,9 β = 13; 8 α ,9 α = 8 α ,9 β = 8 β ,9 β = 4; 8 β ,9 α = 12,12' = 12; OSen: 2,4 = 2,5 = 1.3] (PTLC: CH₂Cl₂-C₆H₆, 1:1, two developments, R_f 0.61).

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